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## Title

### ***Bacterial Polysaccharide-Stabilized Silver Nanoparticles Photocatalytically Decolorize Azo Dyes***

## Abstract

Bacterial polysaccharide is advantageous over plant, algal, and fungal polysaccharides in terms of stability, non-toxicity, and biodegradable nature. In addition, bacterial cell wall polysaccharide (CPs) is very little explored compared to exopolysaccharide. In this study, CPs have been isolated from thermotolerant *Chryseobacterium geocarposphaerae* DD3 (CPs3) from textile industry dye effluent. Structural characterization of the CPs was done by different techniques, viz., scanning electron microscopy-energy dispersive X-ray spectrometry (SEM-EDX), atomic force microscopy (AFM), Fourier transform infrared spectroscopy (FTIR), nuclear magnetic resonance (NMR) spectroscopy, and thermogravimetric analysis (TGA). CPs3 demonstrated compact non-porous amorphous surface composed of evenly distributed macromolecular lumps. TGA revealed a high thermostability (~ 350 °C) of the polysaccharide. FTIR and NMR confirm the polysaccharidic nature of the polymer, consisting of glucose units linked by both  $\beta$ -(1  $\rightarrow$  3) and  $\beta$ -(1  $\rightarrow$  4) glycosidic bonds. The functional properties of CPs3 were evaluated for industrial use as additive, especially antibacterial, emulsification, and flocculation capacities. A single-step green synthesis of silver nanoparticle (AgNP) was performed using CPs3. AgNP was characterized using ultraviolet-visible (UV-Vis) spectroscopy, transmission electron microscopy (TEM), AFM, and particle size analyses. The CPs3-stabilized AgNP exhibited potential photocatalytic activity against a broad range of azo dyes, congo red ( $88.33 \pm 0.48\%$ ), methyl red ( $76.81 \pm 1.03\%$ ), and malachite green ( $47.34 \pm 0.90\%$ ) after only 3 h of reaction. According to our knowledge, this is the first report on CPs from *C. geocarposphaerae*. The results demonstrated multifunctionality of CPs3 in both prospective, CPs3 as additive in

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biotechnology industry as well as Cps3-stabilized AgNP for bioremediation of azo dye. © The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2023.

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## Author Keywords

Azo dye; Cell wall polysaccharide; Chryseobacterium; Photocatalysis; Silver nanoparticle

## Index Keywords

Azo Compounds; Catalysis; Coloring Agents; Metal Nanoparticles; Polysaccharides, Bacterial; Silver; Spectroscopy, Fourier Transform Infrared; Bioremediation; Effluents; Emulsification; Fourier transform infrared spectroscopy; High resolution transmission electron microscopy; Metal complexes; Metal nanoparticles; Nuclear magnetic resonance spectroscopy; Particle size; Particle size analysis; Photocatalytic activity; Scanning electron microscopy; Silver nanoparticles; Synthesis (chemical); Thermogravimetric analysis; X ray powder diffraction; antibiotic agent; azo dye; bacterial polysaccharide; chloramphenicol; congo red; glucose; malachite green; methyl red; penicillin G; polymer; polysaccharide; polysorbate 80; silver nanoparticle; tetracycline; xanthan; azo compound; coloring agent; metal nanoparticle; silver; Atomic-force-microscopy; Azo-dyes; Bacterial cells; Bacterial polysaccharides; Cell-wall-polysaccharides; Chryseobacterium; Exopolysaccharides; Fungal polysaccharides; Non-toxicity; Plant polysaccharides; antibacterial activity; Article; atomic force microscopy; bactericidal activity; bioremediation; biotechnology; cell wall; Chryseobacterium; controlled study; decolorization; effluent; emulsion; energy dispersive X ray spectroscopy; flocculation; Fourier transform infrared spectroscopy; morphology; nonhuman; nuclear magnetic resonance; nuclear magnetic resonance spectroscopy; particle size; photocatalysis; scanning electron microscopy; structure analysis; synthesis; textile industry; thermogravimetry; thermostability; transmission electron microscopy; ultraviolet visible spectroscopy; X ray spectroscopy; zone of inhibition; catalysis; chemistry;

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Azo dyes

## Chemicals/CAS

chloramphenicol, 134-90-7, 2787-09-9, 56-75-7; congo red, 573-58-0, 80701-77-5; glucose, 50-99-7, 84778-64-3, 8027-56-3; malachite green, 569-64-2; methyl red, 493-52-7; penicillin G, 1406-05-9, 61-33-6; polysorbate 80, 8050-83-7, 9005-65-6; tetracycline, 23843-90-5, 60-54-8, 64-75-5, 8021-86-1; xanthan, 11138-66-2; silver, 7440-22-4; Azo Compounds, ; Coloring Agents, ; Polysaccharides, Bacterial, ; Silver,

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## References

Machado T.C., Lansarin M.A., Ribeiro C.S., Wastewater remediation using a spiral shaped reactor for photochemical reduction of hexavalent chromium, *Photochemical & Photobiological Sciences*, 14, 3, pp. 501-505, (2015); Mohan S., Oluwafemi O.S., Kalarikkal N., Thomas S., Songca S.P., *Biopolymers—application in nanoscience and*

---

nanotechnology, Recent advances in biopolymers, 1, 1, pp. 47-66, (2016); Scheffel U., Rhodes B.A., Natarajan T.K., Wagner H.N., Albumin microspheres for study of the reticuloendothelial system, Journal of Nuclear Medicine, 13, 7, pp. 498-503, (1972); Banerjee A., Halder U., Bandopadhyay R., Preparations and applications of polysaccharide based green synthesized metal nanoparticles: A state-of-the-art, Journal of Cluster Science, 28, pp. 1803-1813, (2017); Sathiyarayanan G., Dineshkumar K., Yang Y.H., Microbial exopolysaccharide-mediated synthesis and stabilization of metal nanoparticles, Critical Reviews in Microbiology, 43, 6, pp. 731-752, (2017); Liu J., Qin G., Raveendran P., Ikushima Y., Facile "green" synthesis, characterization, and catalytic function of  $\beta$ -D-glucose-stabilized Au nanocrystals, Chemistry-A European Journal, 12, 8, pp. 2131-2138, (2006); Balantrapu K., Goia D.V., Silver nanoparticles for printable electronics and biological applications, Journal of Materials Research, 24, 9, pp. 2828-2836, (2009); Turner R.D., Mesnage S., Hobbs J.K., Foster S.J., Molecular imaging of glycan chains couples cell-wall polysaccharide architecture to bacterial cell morphology, Nature Communications, 9, 1, (2018); Sundar S., Kundu J., Kundu S.C., Biopolymeric nanoparticles, Science and Technology of Advanced Materials, 11, 1, (2010); Mavaei M., Chahardoli A., Shokoohinia Y., Khoshroo A., Fattahi A., One-step synthesized silver nanoparticles using isoimperatorin: Evaluation of photocatalytic, and electrochemical activities, Scientific Reports, 10, 1, pp. 1762-1774, (2020); Zuurro A., Lavecchia R., Monaco M.M., Iervolino G., Vaiano V., Photocatalytic degradation of azo dye reactive violet 5 on Fe-doped titania catalysts under visible light irradiation, Catalysts, 9, 8, (2019); Sarkar S., Banerjee A., Halder U., Biswas R., Bandopadhyay R., Degradation of synthetic azo dyes of textile industry: A sustainable approach using microbial enzymes, Water Conservation Science and Engineering, 2, pp. 121-131, (2017); Han F., Kambala V.S.R., Dharmarajan R., Liu Y., Naidu R., Photocatalytic degradation of azo dye acid orange 7 using different light sources over Fe<sub>3</sub>+<sup>+</sup>-doped TiO<sub>2</sub> nanocatalysts,

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---

Environmental Technology & Innovation, 12, pp. 27-42, (2018); Samarghandi M.R., Dargahi A., Zolghadr Nasab H., Ghahramani E., Salehi S., Degradation of azo dye acid red 14 (AR14) from aqueous solution using H<sub>2</sub>O<sub>2</sub>/ZVI and S<sub>2</sub>O<sub>8</sub><sup>2-</sup>/ZVI processes in the presence of UV irradiation, Water Environment Research, 92, 8, pp. 1173-1183, (2020); Zhou Y., Tang R.C., Facile and eco-friendly fabrication of colored and bioactive silk materials using silver nanoparticles synthesized by two flavonoids, Polymers, 10, 4, (2018); Sarkar S., Echeverria-Vega A., Banerjee A., Bandopadhyay R., Decolourisation and biodegradation of textile di-azo dye Congo red by *Chryseobacterium geocarposphaerae* DD3, Sustainability, 13, 19, (2021); Sengupta S., Banerjee A., Halder U., Gupta P., Banerjee C., Bandopadhyay R., Comparative study on structure of exopolysaccharide and capsular polysaccharide produced by Southern ocean origin *Pseudoalteromonas* sp. MB-16, Proceedings of the National Academy of Sciences, India Section B: Biological Sciences, 89, pp. 283-290, (2019); Wang Y., Li C., Liu P., Ahmed Z., Xiao P., Bai X., Physical characterization of exopolysaccharide produced by *Lactobacillus plantarum* KF5 isolated from Tibet Kefir, Carbohydrate Polymers, 82, 3, pp. 895-903, (2010); Forato L.A., Bernardes-Filho R., Colnago L.A., Protein structure in KBr pellets by infrared spectroscopy, Analytical Biochemistry, 259, pp. 136-141, (1998); Banerjee A., Das D., Rudra S.G., Mazumder K., Andler R., Bandopadhyay R., Characterization of exopolysaccharide produced by *Pseudomonas* sp. PFAB4 for synthesis of EPS-coated AgNPs with antimicrobial properties, Journal of Polymers and the Environment, 28, pp. 242-256, (2020); Pal P., Banerjee A., Soren K., Chakraborty P., Pandey J.P., Sen G., Bandopadhyay R., Novel biocide based on cationic derivative of Psyllium: Surface modification and antibacterial activity, Journal of Polymers and the Environment, 27, pp. 1178-1190, (2019); Pu L., Zeng Y.J., Xu P., Li F.Z., Zong M.H., Yang J.G., Lou W.Y., Using a novel polysaccharide BM2 produced by *Bacillus megaterium* strain PL8 as an efficient bioflocculant for wastewater treatment,



---

International Journal of Biological Macromolecules, 162, pp. 374-384, (2020); Cooper D.G., Goldenberg B.G., Surface-active agents from two Bacillus species, Applied and Environmental Microbiology, 53, 2, pp. 224-229, (1987); Maity G.N., Mondal S., Mabhai S., Ali S.R., Capsular polysaccharide from Klebsiella Pneumonia ATCC 70063: Isolation, green synthesis of silver nanoparticles and study of antimicrobial activity, Science Journal of Microbiology, (2016); Vanaamudan A., Sadhu M., Pamidimukkala P., Chitosan-Guar gum blend silver nanoparticle bionanocomposite with potential for catalytic degradation of dyes and catalytic reduction of nitrophenol, Journal of Molecular Liquids, 271, pp. 202-208, (2018); Das K.C., Dhar S.S., Rapid catalytic degradation of malachite green by MgFe<sub>2</sub>O<sub>4</sub> nanoparticles in presence of H<sub>2</sub>O<sub>2</sub>, Journal of Alloys and Compounds, 828, (2020); Ahmed Z., Wang Y., Anjum N., Ahmad A., Khan S.T., Characterization of exopolysaccharide produced by Lactobacillus kefiranofaciens ZW3 isolated from Tibet kefir-Part II, Food Hydrocolloids, 30, 1, pp. 343-350, (2013); Kanamarlapudi S.L.R.K., Muddada S., Characterization of exopolysaccharide produced by Streptococcus thermophilus CC30, Biomed Research International, 2017, (2017); Pooja K.P., Chandra T.S., Production and partial characterization of a novel capsular polysaccharide KP-EPS produced by Paenibacillus pabuli strain ATSKP, World Journal of Microbiology and Biotechnology, 25, pp. 835-841, (2009); Sajna K.V., Sukumaran R.K., Gottumukkala L.D., Jayamurthy H., Dhar K.S., Pandey A., Studies on structural and physical characteristics of a novel exopolysaccharide from Pseudozyma sp. NII 08165, International Journal of Biological Macromolecules, 59, pp. 84-89, (2013); Abbas K.A., Abdulkarim S.M., Saleh A.M., Ebrahimian M., Suitability of viscosity measurement methods for liquid food variety and applicability in food industry-A review, Journal of Food Agriculture and Environment, 8, 3-4, pp. 100-107, (2010); de Aguiar K.L.N.P., Palermo L.C.M., Mansur C.R.E., Polymer viscosifier systems with potential application for enhanced oil recovery: A review, Oil & Gas Science and Technology-Revue d'IFP Energies nouvelles, 76, (2021); Cerna M., Barros A.S.,

---

Nunes A., Rocha S.M., Delgadillo I., Copikova J., Coimbra M.A., Use of FT-IR spectroscopy as a tool for the analysis of polysaccharide food additives, *Carbohydrate Polymers*, 51, 4, pp. 383-389, (2003); Chylinska M., Szymanska-Chargot M., Zdunek A., FT-IR and FT-Raman characterization of non-cellulosic polysaccharides fractions isolated from plant cell wall, *Carbohydrate Polymers*, 154, pp. 48-54, (2016); Szymanska-Chargot M., Zdunek A., Use of FT-IR spectra and PCA to the bulk characterization of cell wall residues of fruits and vegetables along a fraction process, *Food Biophysics*, 8, pp. 29-42, (2013); Banerjee A., Mohammed Breig S.J., Gomez A., Sanchez-Arevalo I., Gonzalez-Faune P., Sarkar S., Bandopadhyay R., Vuree S., Cornejo J., Tapia J., Bravo G., Cabrera-Barjas G., Optimization and characterization of a novel exopolysaccharide from *Bacillus haynesii* Camb6 for food applications, *Biomolecules*, 12, 6, (2022); Halder U., Roy R.K., Biswas R., Khan D., Mazumder K., Bandopadhyay R., Synthesis of copper oxide nanoparticles using capsular polymeric substances produced by *Bacillus altitudinis* and investigation of its efficacy to kill pathogenic *Pseudomonas aeruginosa*, *Chemical Engineering Journal Advances*, 11, (2022); Gieroba B., Krysa M., Wojtowicz K., Wiater A., Pleszczyńska M., Tomczyk M., Sroka-Bartnicka A., The FT-IR and Raman spectroscopies as tools for biofilm characterization created by cariogenic streptococci, *International Journal of Molecular Sciences*, 21, 11, (2020); Nagaraj V., Skillman L., Li D., Foreman A., Xie Z., Ho G., Characterisation of extracellular polysaccharides from bacteria isolated from a full-scale desalination plant, *Desalination*, 418, pp. 9-18, (2017); Cui S.W., *Food carbohydrates: Chemistry, physical properties, and applications*, CRC Press, (2005); Cai P., Moran J., Pavliak V., Deng C., Khoury N., Marcq O., Ruppen M.E., NMR structural analysis of the capsular polysaccharide from *Streptococcus pneumoniae* serotype 6C, *Carbohydrate research*, 351, pp. 98-107, (2012); Merino S., Canals R., Knirel Y.A., Tomas J.M., Molecular and chemical analysis of the lipopolysaccharide from *Aeromonas hydrophila* strain AH-1 (Serotype O11), *Marine drugs*, 13, 4, pp. 2233-2249, (2015);

---

---

Karami P., Khorshidi B., McGregor M., Peichel J.T., Soares J.B., Sadrzadeh M., Thermally stable thin film composite polymeric membranes for water treatment: A review, *Journal of Cleaner Production*, 250, (2020); Zhang Y., Wu Y.T., Zheng W., Han X.X., Jiang Y.H., Hu P.L., Tang Z.X., Shi L.E., The antibacterial activity and antibacterial mechanism of a polysaccharide from *Cordyceps cicadae*, *Journal of functional foods*, 38, pp. 273-279, (2017); Qiu H., Si Z., Luo Y., Feng P., Wu X., Hou W., Zhu Y., Chan-Park M.B., Xu L., Huang D., The mechanisms and the applications of antibacterial polymers in surface modification on medical devices, *Frontiers in Bioengineering and Biotechnology*, 8, (2020); Study of pyridinium-type functional polymers. II. Antibacterial activity of soluble pyridinium-type polymers, *Journal of Applied Polymer Science*, 67, 10, pp. 1761-1768, (1998); Banerjee A., Das D., Andler R., Bandopadhyay R., Green synthesis of silver nanoparticles using exopolysaccharides produced by *Bacillus anthracis* PFAB2 and its biocidal property, *Journal of Polymers and the Environment*, 29, pp. 2701-2709, (2021); Luo L., Wu Y., Liu C., Huang L., Zou Y., Shen Y., Lin Q., Designing soluble soybean polysaccharides-based nanoparticles to improve sustained antimicrobial activity of nisin, *Carbohydrate polymers*, 225, (2019); Scala A., Piperno A., Hada A., Astilean S., Vulpoi A., Ginestra G., Marino A., Nostro A., Zammuto V., Gugliandolo C., Marine bacterial exopolymers-mediated green synthesis of noble metal nanoparticles with antimicrobial properties, *Polymers*, 11, 7, (2019); Czuprynski C.J., Noel E.J., Adlam C., Modulation of bovine neutrophil antibacterial activities by *Pasteurella haemolytica* A1 purified capsular polysaccharide, *Microbial Pathogenesis*, 6, 2, pp. 133-141, (1989); Al-Dhabi N.A., Esmail G.A., Arasu M.V., Sustainable conversion of palm juice wastewater into extracellular polysaccharides for absorption of heavy metals from Saudi Arabian wastewater, *Journal of Cleaner Production*, 277, (2020); Roselet F., Vandamme D., Roselet M., Muylaert K., Abreu P.C., Screening of commercial natural and synthetic cationic polymers for flocculation of freshwater and marine microalgae and effects of molecular weight and charge density, *Algal*

---

research, 10, pp. 183-188, (2015); Okaiyeto K., Nwodo U.U., Mabinya L.V., Okoh A.I., Characterization of a bioflocculant produced by a consortium of Halomonas sp. Okoh and Micrococcus sp. Leo, International Journal of Environmental Research and Public Health, 10, 10, pp. 5097-5110, (2013); Mathivanan K., Chandirika J.U., Vinothkanna A., Govindarajan R.K., Meng D., Yin H., Characterization and biotechnological functional activities of exopolysaccharides produced by Lysinibacillus fusiformis KMNTT-10, Journal of Polymers and the Environment, 29, pp. 1742-1751, (2021); Kanmani P., Satish Kumar R., Yuvaraj N., Paari K.A., Pattukumar V., Arul V., Production and purification of a novel exopolysaccharide from lactic acid bacterium Streptococcus phocae PI80 and its functional characteristics activity in vitro, Bioresource technology, 102, 7, pp. 4827-4833, (2011); Bibi A., Xiong Y., Rajoka M.S.R., Mehwish H.M., Radicetti E., Umair M., Shoukat M., Khan M.K.I., Aadil R.M., Recent advances in the production of exopolysaccharide (EPS) from Lactobacillus spp. and its application in the food industry: A review, Sustainability, 13, 22, (2021); Desai R., Mankad V., Gupta S.K., Jha P.K., Size distribution of silver nanoparticles: UV-visible spectroscopic assessment, Nanoscience and Nanotechnology Letters, 4, 1, pp. 30-34, (2012); Darroudi M., Ahmad M.B., Abdullah A.H., Ibrahim N.A., Green synthesis and characterization of gelatin-based and sugar-reduced silver nanoparticles, International Journal of Nanomedicine, pp. 569-574, (2011); Saravanan C., Rajesh R., Kaviarasan T., Muthukumar K., Kavitate D., Shetty P.H., Synthesis of silver nanoparticles using bacterial exopolysaccharide and its application for degradation of azo-dyes, Biotechnology Reports, 15, pp. 33-40, (2017); Santipanichwong R., Suphantharika M., Weiss J., McClements D.J., Core-shell biopolymer nanoparticles produced by electrostatic deposition of beet pectin onto heat-denatured  $\beta$ -lactoglobulin aggregates, Journal of Food Science, 73, 6, pp. N23-N30, (2008); Raza Z.A., Noor S., Majeed M.I., PEGylation of poly (hydroxybutyrate) into multicomponent nanostructures and loading thereon with bioactive molecules for potential biomedical applications, Journal of Polymer

---

Research, 28, pp. 1-11, (2021); Eiden-Assmann S., Widoniak J., Maret G., Synthesis and characterization of porous and nonporous monodisperse colloidal TiO<sub>2</sub> particles, Chemistry of Materials, 16, 1, pp. 6-11, (2004); Zielinska A., Carreiro F., Oliveira A.M., Neves A., Pires B., Venkatesh D.N., Durazzo A., Lucarini M., Eder P., Silva A.M., Santini A., Souto E.B., Polymeric nanoparticles: Production, characterization, toxicology and ecotoxicology, Molecules, 25, 16, (2020); Bryers J.D., Mason C.A., Biopolymer particulate turnover in biological waste treatment systems: A review, Bioprocess Engineering, 2, pp. 95-109, (1987); Li J., Liu Y., Zhu Z., Zhang G., Zou T., Zou Z., Zhang S., Zeng D., Xie C., A full-sunlight-driven photocatalyst with super long-persistent energy storage ability, Scientific reports, 3, 1, (2013); Asghar A., Raman A.A.A., Daud W.M.A.W., Advanced oxidation processes for in-situ production of hydrogen peroxide/hydroxyl radical for textile wastewater treatment: A review, Journal of Cleaner Production, 87, pp. 826-838, (2015); Guan R., Yuan X., Wu Z., Jiang L., Li Y., Zeng G., Principle and application of hydrogen peroxide based advanced oxidation processes in activated sludge treatment: A review, Chemical Engineering Journal, 339, pp. 519-530, (2018); Rajkumar R., Ezhumalai G., Gnanadesigan M., A green approach for the synthesis of silver nanoparticles by *Chlorella vulgaris* and its application in photocatalytic dye degradation activity, Environmental Technology & Innovation, 21, (2021); Mehwish H.M., Rajoka M.S.R., Xiong Y., Cai H., Aadil R.M., Mahmood Q., He Z., Zhu Q., Green synthesis of a silver nanoparticle using *Moringa oleifera* seed and its applications for antimicrobial and sunlight mediated photocatalytic water detoxification, Journal of Environmental Chemical Engineering, 9, 4, (2021); Ahmed T., Noman M., Shahid M., Niazi M.B.K., Hussain S., Manzoor N., Wang X., Li B., Green synthesis of silver nanoparticles transformed synthetic textile dye into less toxic intermediate molecules through LC-MS analysis and treated the actual wastewater, Environmental Research, 191, (2020); Mangalam J., Kumar M., Sharma M., Joshi M., High adsorptivity and visible light assisted photocatalytic activity of silver/reduced graphene oxide (Ag/rGO)

---

nanocomposite for wastewater treatment, *Nano-Structures & Nano-Objects*, 17, pp. 58-66, (2019); Roushani M., Mavaei M., Rajabi H.R., Graphene quantum dots as novel and green nano-materials for the visible-light-driven photocatalytic degradation of cationic dye, *Journal of Molecular Catalysis A: Chemical*, 409, pp. 102-109, (2015); Silva Z.S., Huang Y.Y., de Freitas L.F., Franca C.M., Botta S.B., Ana P.A., Mesquita-Ferrari R.A., Fernandes K.P.S., Deana A., Leal C.R.L., Prates R.A., Hamblin M.R., Bussadori S.K., Papain gel containing methylene blue for simultaneous caries removal and antimicrobial photoinactivation against *Streptococcus mutans* biofilms, *Scientific reports*, 6, 1, pp. 1-12, (2016); Silva I.M.P., Byzinski G., Ribeiro C., Longo E., Different dye degradation mechanisms for ZnO and ZnO doped with N (ZnO: N), *Journal of Molecular Catalysis A: Chemical*, 417, pp. 89-100, (2016); Cittrarasu V., Kaliannan D., Dharman K., Maluventhen V., Easwaran M., Liu W.C., Balasubramanian B., Arumugam M., Green synthesis of selenium nanoparticles mediated from *Ceropegia bulbosa* Roxb extract and its cytotoxicity, antimicrobial, mosquitocidal and photocatalytic activities, *Scientific reports*, 11, 1, (2021); Wu Z.C., Zhang Y., Tao T.X., Zhang L., Fong H., Silver nanoparticles on amidoxime fibers for photo-catalytic degradation of organic dyes in waste water, *Applied Surface Science*, 257, 3, pp. 1092-1097, (2010)

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